Whole Genome Sequencing, Assembly and Annotation

Strategy

Libraries

Sequencing

Assembly

Closure

Annotation

Release

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Introduction

- Whole genome sequencing provide information about coding and noncoding part of genome.
- To fetch out important pathways.
- For evolutionary studies and species comparison.
- I For more effective personalized medicine (why a drug works for person X and not for Y).
- Disease-susceptibility prediction based on gene sequence variation.

History of Sequencing

- Allan Maxam and Walter Gilbert developed an important method of DNA sequencing in 1976-1977.
- This method of chemical modification of DNA was technically complex and fallen out of flavor due to the use of extensive hazardous chemicals, and difficulties with scale-up.

History of Sequencing

- Sanger and his team developed the chaintermination method of DNA sequencing in 1977.
- Only be used for fairly short strands (100 to 1000 base pairs) and longer sequences must be subdivided into smaller fragments.
- After this, these small fragments subsequently reassembled to give the overall sequence

History of Sequencing

- Shotgun sequencing has been developed for sequencing of large fragments of DNA in 1979.
- DNA is broken up randomly into numerous small segments, which are sequenced using the chain termination method and then short reads have been produced.
- Shotgun sequencing was the initiative for full genome sequencing.

WHOLE GENOME SEQUENCING

- Information about coding and non coding part of an organism.
- To find out important pathways in microbes.
- For evolutionary study and species comparison.
- For more effective personalized medicine (why a drug works for person X and not for Y).
- Identification of important secondary metabolite pathways (e.g. in plants).
- Disease-susceptibility prediction based on gene sequence variation.

NEXT GENERATION SEQUENCING

- Sequence full genome of an organism in a few days at a very low cost.
- Produce high throughput data in form of short reads.



Illumin a



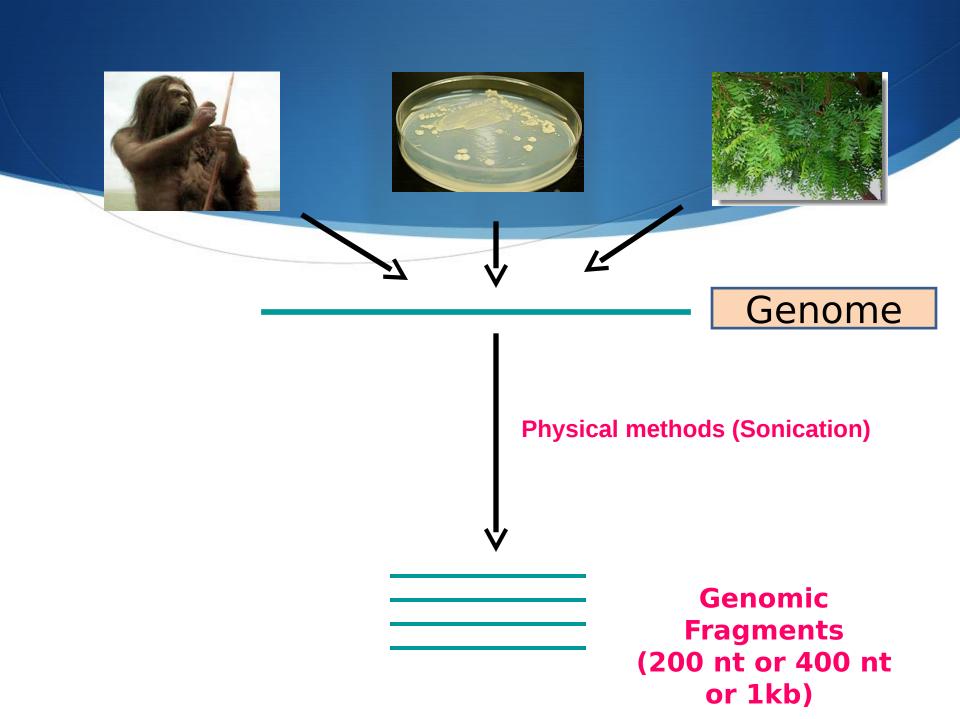
ABI's Solid

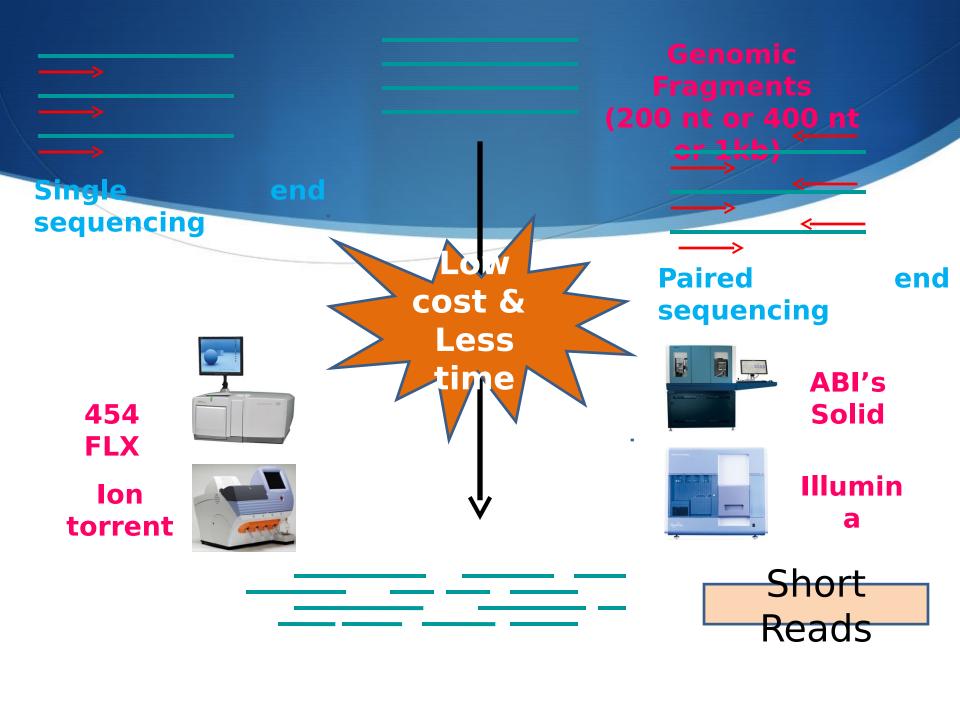


Roche's 454 FLX



lon torrent





CHALLENGES

Removal of artifacts in short reads ??

Genome assembly of short reads ??



Several assemblers available, which is best ??

Annotation and validation of assembled genome ??

Recent techniques

- High throughput sequencing also called Next Generation Sequencing (NGS) have the capacity to sequence full genomes.
- These technologies Includes Roche's 454 GS FLX, Illumina's Solexa technology, ABI's SOLiD technology and Ion torrent technology.

Next Generation Sequencing

Technique	lon torrent	Roche's 454	Illumina	ABI's SOLiD
Data (Mb per run)	100	100	600	700
Time per run	1.5 Hrs	7 Hrs	9 Days	9 Days
Read length	200 bp	400 bp	150 bp	75 bp
Cost per Mb	5 \$	84.39 \$	0.03 \$	0.04 \$

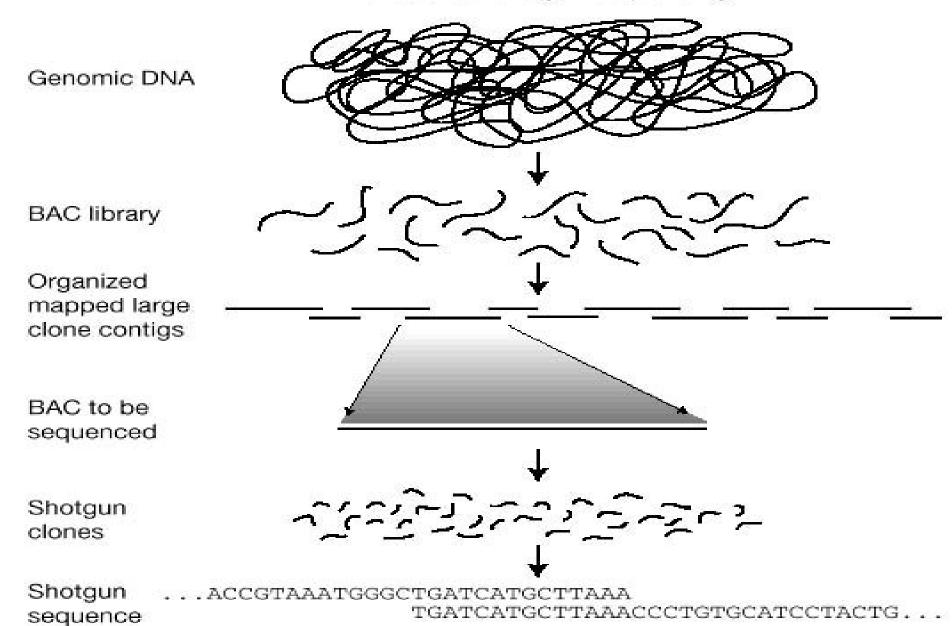
History of genome sequencing

- Bacteriophage fX174, was the first genome to be sequenced, a viral genome with only 5,368 base pairs (bp).
- First bacterial genome sequenced was Haemophilus influenza.
- The first nearly complete human genomes sequenced were J. Craig Venter's, James Watson's, a Han Chinese, a Yoruban from Nigeria, a female leukemia patient, and Seong-Jin Kim.
- As of June 2012, there are 69 nearly complete human genomes publicly available.

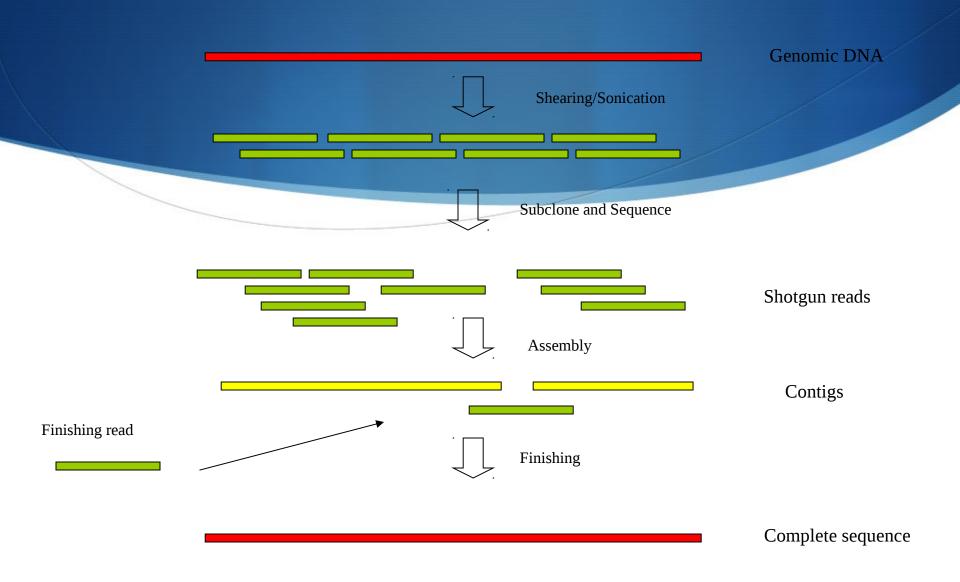
Challenges of genome sequencing

- Data produce in form of short reads, which have to be assembled correctly in large contigs and chromosomes.
- Short reads produced have low quality bases and vector/adaptor contaminations.
- Several genome assemblers are available but we have to check the performance of them to search for best one.

Hierarchical shotgun sequencing



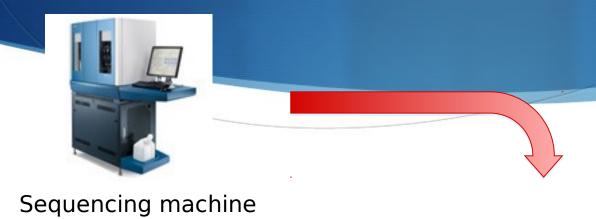
Assembly ... ACCGTAAATGGGCTGATCATGCTTAAACCCTGTGCATCCTACTG...



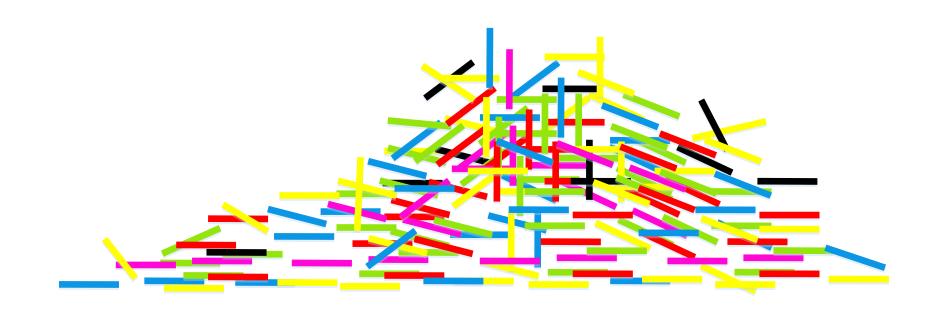
Short read alignment



Short read alignment



And you get MANY of them



De novo assembly strategies

SSAKE

- Warren et al., 2007
- Uses DNA prefix tree to find k-mer matches

Edena

- Hernandez et al., 2008
 - overlap-layout algorithm adapted for short reads

Velvet

- Zerbino and Birney, 2008
- Uses DeBruijn graph algorithm plus error correction

Comparative assembly using multiple genomes

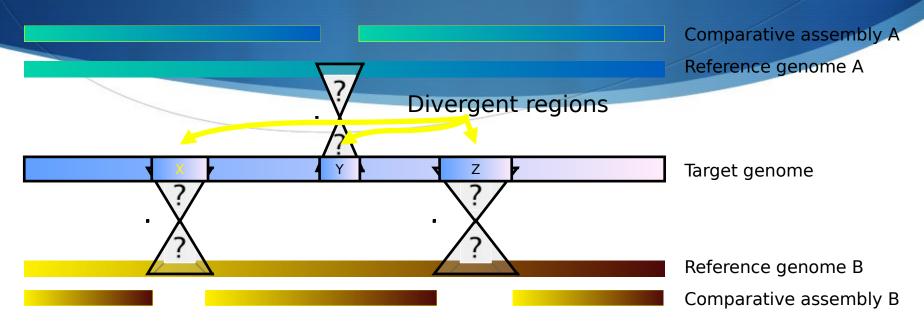
Assembly A

Assembly B

Merge

Merged assembly

Comparative assembly using multiple genomes



Genome annotation

- A process of attaching biological information to sequences (contigs or chromosomes).
- Consists of two main steps: -
- A. Identifying elements on genome a process called gene prediction (Structural annotation).
- B. Attaching biological information to these elements (Functional annotation).

Genome annotation

- Structural annotation
- ORFs and their localisation
- Gene structure
- Coding regions
- Location of regulatory motifs
- Functional annotation
- Biochemical function
- Biological function
- Involved regulation and interactions
- Expression

Genome annotation

- Can be done manually (require human expertise) or with automated pipelines.
- Pipelines available :-
- > PGAAP (NCBI)
- > RAST server
- > IMG-ER,
- > ISGA
- MAKER (for eukaryotes).

Genome annotation tools at IMTECH

- Protein Structure prediction servers
- Servers for predicting function of proteins
- Servers for designing epitope based vaccine
- Genome annotation
- Molecular Interactions & Modifications
- Designing of Therapeutic Molecules
- Computer Aided Drug Design

http://www.imtech.res.in/raghava/

Genome submission to NCBI (GenBank)

- NCBI (GenBank) accepts both complete and incomplete genomes (contigs produced after genome assembly).
- Bacterial genome submission instructions available at http://www.ncbi.nlm.nih.gov/genbank/genomesubmit/.
- Eukaryotic genome submission instructions availble at
- http://www.ncbi.nlm.nih.gov/genbank/eukaryotic genome submission/

Publications

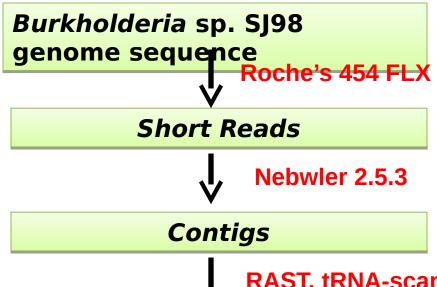
- Whole genome assembly and annotation of microbes with preliminary analysis can be published in reputed journals like Journal of Bacteriology (http://jb.asm.org/) and Eukaryotic cell (http://jb.asm.org/).
- Other journals are Genome Biology, Genome Reaserch and Nature Biotechnology(according to the analysis done).

Genome assembly and annotation done at IMTECH

- Burkholderia sp. SJ98 (Kumar et al. 2012).
- Debaryomyces hansenii MTCC 234 (Kumar et al. 2012).
- Imtechella halotolerans K1^T (Kumar et al. 2012).
- Marinilabilia salmonicolor JCM 21150[⊤] (Kumar et al. 2012).
- Rhodococcus imtechensis sp. RKJ300 (Vikram et al. 2012).
- Rhodosporidium toruloides MTCC 457 (Kumar et al. 2012).

Burkholderia sp. SJ98

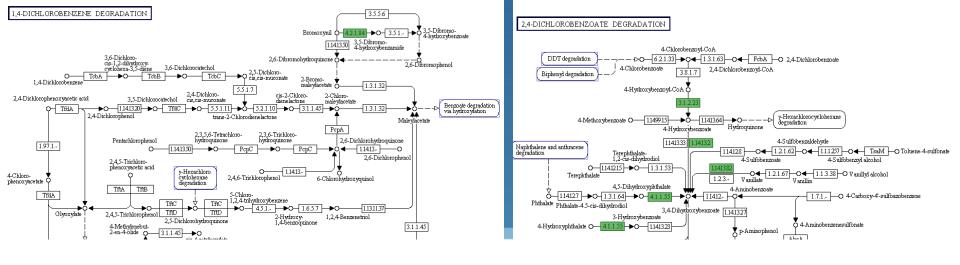
Degrade a number of aromatic compounds, e.g., p nitrophenol, o-nitrobenzoate, p-nitrobenzoate, and 4-nitrocatechol (Pandey G, et. al. 2002), 2-chloro-4-nitrophenol (Pandey J, et al. 2011), and 3-methyl-4-nitrophenol (Bhushan B, et. al. 2000).



Genome size	7.89-Mb
Large contigs	79
Protein coding genes	7,364
rRNAs	3
tRNAs	51

RAST, tRNA-scan v1.21 and RNAmmer v1.2

Annotated genome

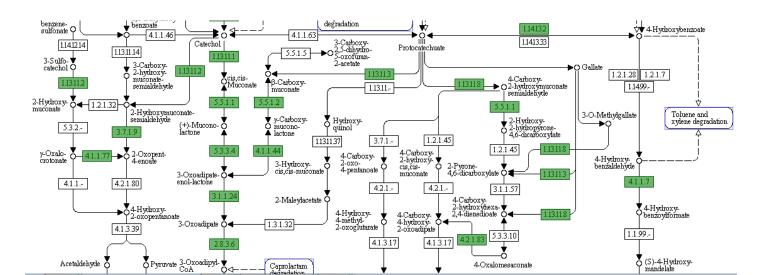


Journal of Bacteriology

Genome Sequence of the Nitroaromatic Compound-Degrading Bacterium Burkholderia sp. Strain SJ98

Shailesh Kumar, Surendra Vikram and Gajendra Pal Singh Raghava

J. Bacteriol. 2012, 194(12):3286. DOI: 10.1128/JB.00497-12.



Azadirachta indica (Neem) Genome and transcriptome assembly and annotation

Dr. Prof. Siddhartha Roy (Director), IICB, Kolkata

Dr. Rupak K. bhadra , IICB, Kolkata

Dr. G P S Raghava, IMTECH, Chandigarh

Dr. Saikat Chakrabarti, IICB, Kolkata

Dr. Prabodh Trivedi, NBRI, Lucknow

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Dr. Mehar Asif, NBRI, Lucknow

Dr. Sridhar Sivasubbu, IGIB, New Delhi,

Dr. Vinod Scaria, IGIB, New Delhi

Azadirachta indica

(Neem)
Each part of the neem tree has some medicinal property and is thus commercially exploitable.

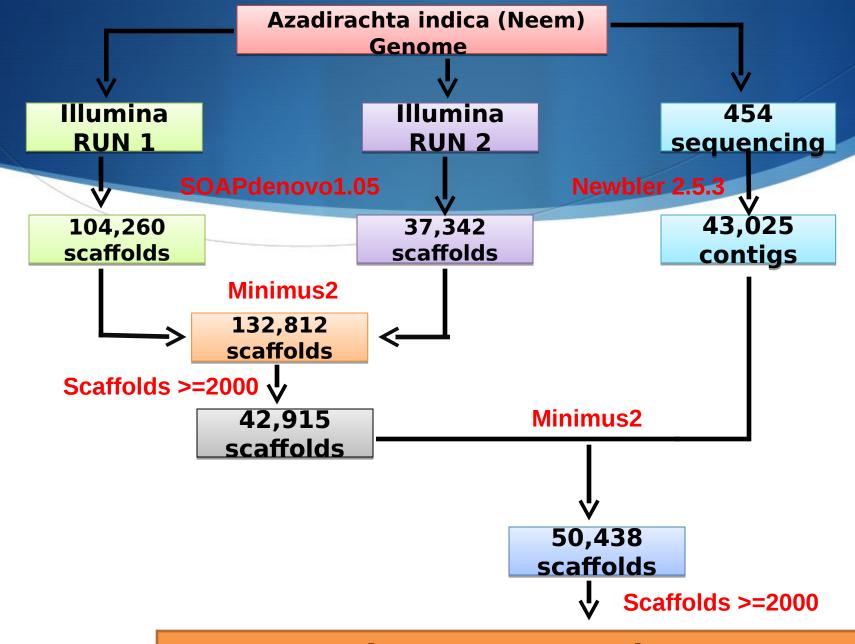


Neem Genome and transcriptome

CAGUANCING			
Genome sequecing	Illumina and Roche's 454		
ranscriptome sequencing	Roche's 454		
Genome assembly	SOAPdenovo and Newbler		
Transcriptome assembly	Newbler		
Gene Prediction	FGENESH and Augustus		
Annotation	BLAST2GO and manualy		
Repeatmasking	Repeatmasking		

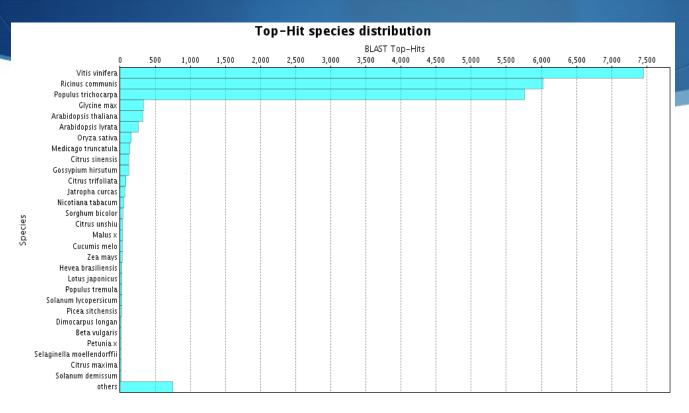
Transcripts mapping to Genome

BLAST programe



Size = 316.304Mb

BLAST2GO annotation



1



Vitis vinifera 487Mb



Ricinu

Populus trichocarpa 485Mb

Ricinus communis 352Mb





Rhodococcus imtechensis RJ300

Strain RKJ300 is capable of utilizing 4 nitrophenol, 2-chloro-4-nitrophenol, and 2, 4-dinitrophenol as sole sources of carbon and energy(Ghosh A, et al. 2010).

Rhodococcus imtechensis sp. RKJ300 | Illumina GAIIX

Short Reads



Filtered Short
Reads
SOAPdenovo v1.05

Genome size	8.231-Mb
Contigs produced	178
Protein coding genes	8,059
rRNAs	5
tRNAs	49

Contigs



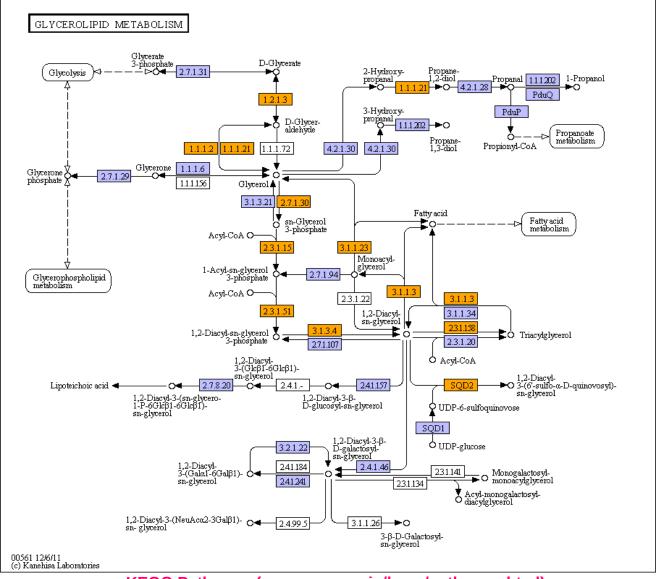
RAST, tRNA-scan v1.21 and RNAmmer v1.2

Annotated genome

Rhodosporidium toruloides MTCC 457

It can accumulate lipids to a higher level (~75% of dry weight under certain conditions) than most other oleaginous yeasts and fungi (Ageitos, J. M. et. al.).

R. toruloides offers many opportunities for being developed as an additional yeast model and synthetic biology platform to *Saccharomyces cerevisiae*.



KEGG Pathways (www.genome.jp/kegg/pathways.html)

Kumar, S., Kushwaha, H., Bachawat, A.K., Raghava G.P.S. and Ganesan, K. Genome sequence of the oleaginous red yeast Rhodosporidium toruloides MTCC 457. Eukaryotic Cell (In Press).

Imtechella halotolerans K1[™]

Strain K1T is known to possess various enzymatic activities, such as lipase, Y-glutamyl transferase, glycine arylamidase, and Glu-Gly-Arg-arylamidase (Vikram S et. al. 2012).

Imtechella halotolerans K1^T



Illumina Hi-Seq 1000

Short Reads



NGS QC toolkit v2.2.1

Filtered Short Reads



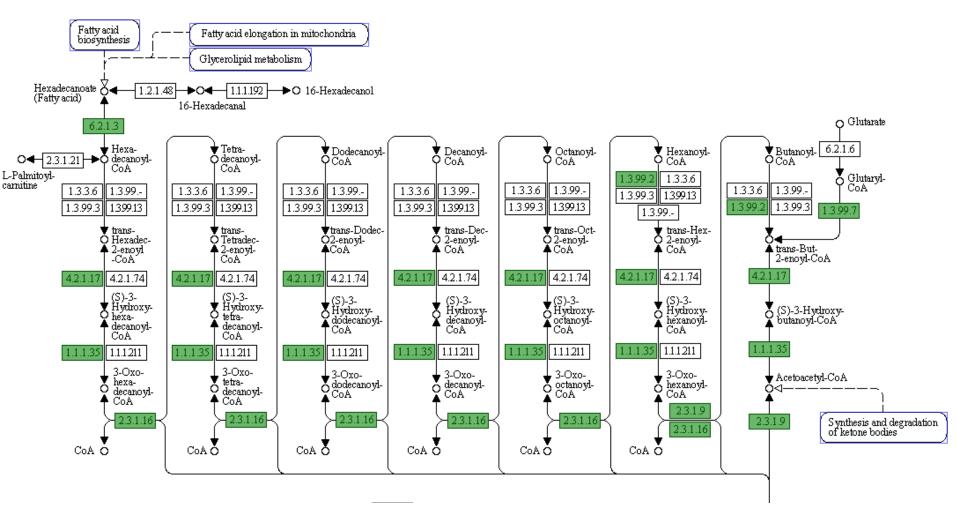
Velvet v1.2.03

Contigs

Genome size	3.087-Mb
Contigs produced	44
Protein coding genes	2,738
rRNAs	3
tRNAs	38

 \checkmark RAST, tRNA-scan v1.21 and RNAmmer v1.2

Annotated genome



Journal of Bacteriology

Genome Sequence of the Halotolerant Bacterium Imtechella halotolerans K1

Shailesh Kumar, Surendra Vikram, Srikrishna Subramanian, Gajendra Pal Singh Raghava and Anil Kumar Pinnaka *J. Bacteriol.* 2012, 194(14):3731. DOI: 10.1128/JB.00506-12.

Marinilabilia salmonicolor JCM21150[™]

The strain is capable of gelatin liquefaction. All the strains of the genus Marinilabilia were reported to decompose various biomacromolecules (Muller HE et. al. 1996).

Marinilabilia sali JCM 21150 [™]			
V	Illumina Hi-Seq 1000		
Short R	eads		
\	NGS QC toolkit v2.2.1		
Filtered Short Reads			
\	Velvet v1.2.03		
Conti	gs		

Genome size	4.98-Mb
Contigs produced	72
Protein coding genes	4,227
rRNAs	3
tRNAs	52
Closest neighbor	Bacteroides sp. 2_1_7



RAST, tRNA-scan v1.21 and RNAmmer v1.2

Annotated genome

Debaryomyces hansenii var. hansenii MTCC234

- D. hansenii is considered a sodium includer, and the accumulation of a large amount of NaCl does not have any adverse effect on its physiology (Prista C. et. al. 2005).
- Besides xylitol, strains of *D. hansenii* are also known to produce arabitol and riboflavin (Breuer U et. al. 2006).
- Compared to *D. hansenii strain CBS767*, whose genome was sequenced previously,MTCC 234 is more halotolerant and it also produces riboflavin and arabitol.

Genomics web portal

Rhodosporodium toruloides

MTCC 457

HOME	Help	Acknowledgement	Developers	Contact
Genomics at BIC (IN	итесн)	Но	me Page	
Genome sequencing Genome assembly	:			
Genome annotation		This is a web portal for all genomics work held at (IMTECH), Chandigarh.	Bioinformatics center of Institute of I	Microbial Technology
Prokaryotes		We have sequenced, assembled and annotate several m	icrobial genomes.	
Actinoalloteichus sp RMV-1378 ^T Burkholderia sp. SJ Rhodococcus rhodoc BKS6-46 Intechella halotoler RkJ300	98 chrous rans Kl ^T	1. Actinoalloteichus spitiensis RMV-1378 ^T 2. Rhodococcus rhodochrous BKS6-46 3. Burkholderia sp. S J 98 4. Imtechella halotolerans K1 ^T 5. Marinilabilia salmonicolor JCM 21150 6. Rhodococcus imtechensis sp. RKJ300 7. Debaryomyces hansenii MTCC 345 8. Rhodosporodium toruloides MTCC 457		
Marinilabilia salmor 21150 Eukaryotes Debaryomyces hans 234		http://crdd.osdd.net/	′raghava/genon	nesrs

CRAG: Computational Resources for Assembly and Annotation of Genomes

G.P.S. Raghava | Bioinformatics Centre | IMTECH | CRDD | Team Members | Contact | FAQ

Bioinformatics

Home About CRAG Infrastructure

Facility to Community
Our Servers

Protocols Panda Genome

References

Sequencing Tech.
Sanger Sequencing

SRS by HTS
Types of Data
Resources

Assemblers for

Long sequences Short Read Seq. Hybrid Seq. Softwares Used Genome Viewers

Annotation

Challenges

Contigs joining Prokaryotic Genomes Eukaryotic Genomes

Important Links

Computation Resources for Assembly and Annotation of Genomes(CRAG)

You are welcome to visit Computation Resources for Assembly and Annotation of Genomes(CRAG) site at Institute of Microbial Technology (IMTECH), Chandigarh. Aim of this site is to assist the users in assembling of genomes from short read sequencing (SRS). Their is exponential growth in SRS data, due to high throughput sequencing (HTS) techniques. We have following major objective

- · Collection and compilation of computation resources
- · Brief Description of genome assemblers
- · Maintaing SRS and related data
- · Service to community to assemble their genomes
- · Analysis of assembled genome
- · Genome Annotation

We are also planning to provide annotation service to scientific community, in addition to genome assembling. Our aim is to provide free service to community using available public domain software.

Vikram S, Kumar S and Raghava GPS, *Denovo* genome assembly and annotation of microbes. OSCAT 2012,IMTECH,Chandigarh (Poster)

http://imtech.res.in/raghava/crag

Mance