



Research report

GLP-1 analogue CJC-1131 prevents amyloid β protein-induced impairments of spatial memory and synaptic plasticity in rats

Sheng-Xiao Zhang^{a,1}, Hong-Yan Cai^{b,1}, Xiao-Wen Ma^a, Li Yuan^c, Jun Zhang^c,
Zhao-Jun Wang^c, Yu-Feng Li^a, Jin-Shun Qi^{c,*}

^a Department of Rheumatology, The Second Hospital of Shanxi Medical University, 56 Xinjian South Road, Taiyuan, Shanxi 030001, China

^b Department of Microbiology and Immunology, Shanxi Medical University, 56 Xinjian South Road, Taiyuan, Shanxi 030001, China

^c Department of Physiology and National Key Discipline of Physiology, Shanxi Medical University, 56 Xinjian South Road, Taiyuan, Shanxi 030001, China

ARTICLE INFO

Article history:

Received 21 November 2016

Received in revised form 4 March 2017

Accepted 8 March 2017

Available online 15 March 2017

Keywords:

CJC-1131

Amyloid β protein

Spatial memory

Morris water maze

Long-term potentiation

Synaptic plasticity

ABSTRACT

Although amyloid β protein ($A\beta$) has been recognized as one of the main pathological characteristics in the brain of Alzheimer's disease (AD), the effective strategies against $A\beta$ neurotoxicity are still deficient up to now. Glucagon-like peptide 1 (GLP-1), a natural gut hormone, was found to be effective in modulating insulin signaling and neural protection, but short half-life limited its clinical application in AD treatment. CJC-1131, a newly designed GLP-1 analogue with very longer half-life, has shown good effectiveness in the treatment of type 2 diabetes mellitus (T2DM). However, it is unclear whether CJC-1131 could alleviate $A\beta$ -induced neurotoxicity in cognitive behavior and electrophysiological property. The present study investigated the effects of CJC-1131 on the $A\beta$ -induced impairments in spatial memory and synaptic plasticity of rats by using Morris water maze test and in vivo field potential recording. The results showed that $A\beta$ 1-42-induced increase in the escape latency of rats in hidden platform test and decrease in swimming time percent in target quadrant were effectively reversed by CJC-1131 pretreatment. Further, CJC-1131 prevented against $A\beta$ 1-42-induced suppression of hippocampal long term potentiation (LTP). In addition, $A\beta$ 1-42 injection resulted in a significant decrease of p-PKA in the hippocampus, which was effectively prevented by CJC-1131 treatment. These results indicated that CJC-1131 protected the cognitive function and synaptic plasticity of rats against $A\beta$ -induced impairments, suggesting that GLP-1 analogue CJC-1131 might be potentially beneficial to the prevention and treatment of AD, especially those with T2DM or blood glucose abnormality.

© 2017 Elsevier B.V. All rights reserved.

1. Introduction

Alzheimer's disease (AD), the leading cause of the senile dementia in the world, is a chronic, incurable neurodegenerative disease, with gradually decayed cognitive function and impaired learning and memory. One of the main pathological characteristics of AD is the accumulation of senile plaques in the brain with AD. Senile plaques are mainly composed of amyloid beta protein ($A\beta$), primarily responsible for the neuronal death [1,2]. It has been universally reported that not only natural amyloid beta proteins such as $A\beta$ 1-42 [3], $A\beta$ 1-40 [4] but also synthetic amyloid beta proteins like

$A\beta$ 25-35 [5] and even $A\beta$ 31-35 [6] can injure neurons in vivo and in vitro. Unluckily, there is no effective strategy to antagonize the neurotoxicity of $A\beta$ up to now.

It is interesting that AD is closely related to type 2 diabetes mellitus (T2DM) in many ways. For example, the prevalence of both diseases (AD and T2DM) was increased exponentially with aging; T2DM raised 1.5–2.5-fold risk of dementia [7,8], while AD patients often have blood glucose abnormalities. The pathological characteristics of both diseases also have much in common. The loss and dysfunction of β -cell in T2DM is related to amyloid deposition [9], which is pretty similar to the aggregated $A\beta$ in the AD brain. Not only $A\beta$, but also hyperphosphorylated tau protein appeared in both AD and T2DM [10]. Moreover, streptozotocin (STZ), used to damage β cells of the pancreas and produce diabetes, also impaired cognition and led to AD model [11]. Schwartz [12] found that the decrease of glucose utilization and the impairment of energy metabolism in AD patients might be related to the damage of the insulin signaling pathway. All of these facts indicate a close

* Corresponding author at: Department of Physiology and Neurobiology Key Laboratory of Cellular Physiology, Ministry of Education, Shanxi Medical University, Taiyuan, Shanxi, 030001, China.

E-mail address: jinshunqi2009@163.com (J.-S. Qi).

¹ With the same contribution.

association between AD and T2DM. Therefore, AD has been considered or named as type 3 diabetes [13]. Importantly, some insulin signal normalizing gut hormones used to treat T2DM such as glucagon-like peptide 1 (GLP-1) have been found to be effective in preventing AD progress. However, natural GLP-1 has a very short half-life of about 2 min [14], due to the rapid degradation by enzyme dipeptidyl peptidase IV (DPP IV), which seriously limits the application of GLP-1 in the clinic.

CJC-1131, a new chemical-modified GLP-1 analogue containing a D-alanine substitution and a maleimidopropylamido attachment to albumin [15], was found to be effective in resisting DPP IV degradation and has a very longer half-life up to 221–353 h [16]. Except for a high dose-related gastrointestinal complaint, no obvious side effect of CJC-1131 was observed in clinical application [17]. The benefits of CJC-1131 in T2DM treatment have been praised, which significantly reduced HbA1c and fasting blood glucose level, improved glucose tolerance and reduced body weight without hypoglycemia adverse effects [17,18]. However, the neuroprotective effects of CJC-1131, as a new GLP-1 analogue with longer half-life, have not been examined up to now. Therefore, the present study explored the neuroprotective effects of intra-hippocampal CJC-1131 injection against A β 1–42-induced impairments in spatial learning and memory of rats. Furthermore, we recorded in vivo hippocampal long-term potentiation (LTP) in the CA1 region of rats to clarify possible association between behavioral changes and electrophysiological properties. Finally, the level of p-PKA in the hippocampus was examined to illuminate the possible molecular mechanism of CJC-1131 in neuroprotection.

2. Materials and methods

2.1. Animals and drugs

Forty adult male Sprague-Dawley (SD) rats (180–220 g) were provided by the Animal Research Center of Shanxi Medical University. All animal procedures were carried out according to the guidelines of Shanxi Committee on Ethics of Animal Research. The rats were kept in a room with 12 h light and 12 h dark cycle at the temperature of $25 \pm 2^\circ\text{C}$ and humidity of 60%–80%. A β 1–42 was ordered from Sigma (Sigma-Aldrich, Germany) and CJC-1131 was synthesized by Sangon (Sangon-Biotech, Shanghai, China). Both of them were dissolved in saline (2.5 nmol/ μl) and stored at -20°C .

2.2. Surgery and intra-hippocampal injection

All rats were randomly divided into four groups: vehicle + saline, A β 1–42 + saline, vehicle + CJC-1131, and A β 1–42 + CJC-1131. To minimize suffering, all surgery was performed under the anesthesia by chloral hydrate intra-peritoneal injection (0.3 g/kg). The SD rat was placed in a stereotaxic apparatus (Narishige, Tokyo, Japan). Drugs were administered into the bilateral hippocampi at the following coordinates [19–21]: anterior/posterior: -3.0 mm , lateral/medial: $\pm 2.2\text{ mm}$, dorsal/ventral: -3.0 mm , with an injection rate of $0.2\text{ }\mu\text{l}/\text{min}$ under the control of a micro pump (RWD Life Science Ltd., Shenzhen, China). CJC-1131 (2 μl) or saline (2 μl) was firstly injected into the bilateral hippocampi. 15 min later, A β 1–42 (2 μl) or vehicle (2 μl) was applied. To make sure that drugs were fully dispersed into the hippocampus, a 5 min-retention of the injection syringe in the brain was given after every injection.

2.3. Morris water maze test

Morris water maze (MWM) test was performed two weeks after surgery [20,22,23]. The maze (Zhenghua Bio Instrument Ltd., China) was a stainless pool (150 cm in diameter and 50 cm in height) with tap water kept at $25 \pm 2^\circ\text{C}$. An escape platform (14 cm in diameter,

29 cm in height) was located in the center of one quadrant and was below the surface of the water about 1 cm. The movement of rat was monitored and recorded by camera above the maze and the signals were analyzed by behavioral analysis software (Noldus Information Technology, Ethovision 3.0, Wageningen, Netherlands). The escape latency and distance to find the escape platform, the percentages of time and distance in target quadrant, and the swimming speed were calculated. In the hidden platform test, rat was placed into the water facing the wall of the pool at a random quadrant. If it didn't find the hidden platform in 120 s, the rat was led to the platform and stayed on the platform for 10 s. Rats were trained four times per day for 5 days. In the probe trial on the sixth day, the platform was removed and the rat was allowed to swim in the pool freely for 120 s. After that, visible platform test was performed, in which the platform was elevated above the water surface about 1 cm.

2.4. In vivo hippocampal LTP recording

After the MWM test, SD rats were anesthetized (urethane, 1.5 g/kg) and placed in a stereotaxic apparatus. An electric blanket was used under body to maintain the rats' temperature at $37\text{--}38^\circ\text{C}$. The skull of rat was drilled, and a bound stimulating/recording electrode was inserted into the Schaffer collateral/CA1 region. Continuous single stimuli with an interval of 30 s were applied to Schaffer to induce field excitatory postsynaptic potentials (fEPSPs) in the CA1 region. Basic fEPSPs were firstly recorded for 30 min as control. Two paired-pulse stimuli were given to trigger paired-pulse facilitation (PPF), and high-frequency stimulation (HFS) with three bursts were given to induce LTP. Each burst had 20 pulses. LTP was recorded for at least 1 h after HFS [19,24].

2.5. Western blot assay

After the electrophysiological experiment, the hippocampi of rats were instantly dissected and frozen at -80°C . Western blot was performed as previously described [19]. Protein concentration was detected with bicinchoninic acid protein assay kit. Total protein was separated by 12% SDS-PAGE and transferred to PVDF membrane. Nonspecific binding was blocked with 5% BSA in Tris-buffered saline containing 0.05% Tween-20 (TBST). Target proteins were detected by GAPDH anti-rat monoclonal antibody (dilution 1:1000; ZSGB-BIO, Beijing, China), phosphorylated PKA anti-rat monoclonal antibody (dilution 1:1000; Abcam, Cambridge, MA, USA) and PKA anti-rat monoclonal antibody (dilution 1:1000; Abcam, Cambridge, MA, USA). Secondary antibody of anti-rabbit immunoglobulin G coupled to horseradish peroxidase (dilution 1:5000) was obtained from ZSGB-BIO. The membrane was rinsed with TBST and the immunocomplex was visualized by using an enhanced chemiluminescence detection kit (Beyotime, Inc). The signals of the membrane were scanned with the FluorChem Scanner and quantified with the Alpha View SA software.

2.6. Data analysis

All data were represented as the mean $\pm$ SEM. For the escape latency in the hidden platform test, three-way repeated measures analysis of variance was used. Other data were analyzed by two-way repeated measures analysis of variance. The software for the statistics was SPSS 13.0 and SigmaPlot 12.3. $p < 0.05$ was taken as statistical significance.

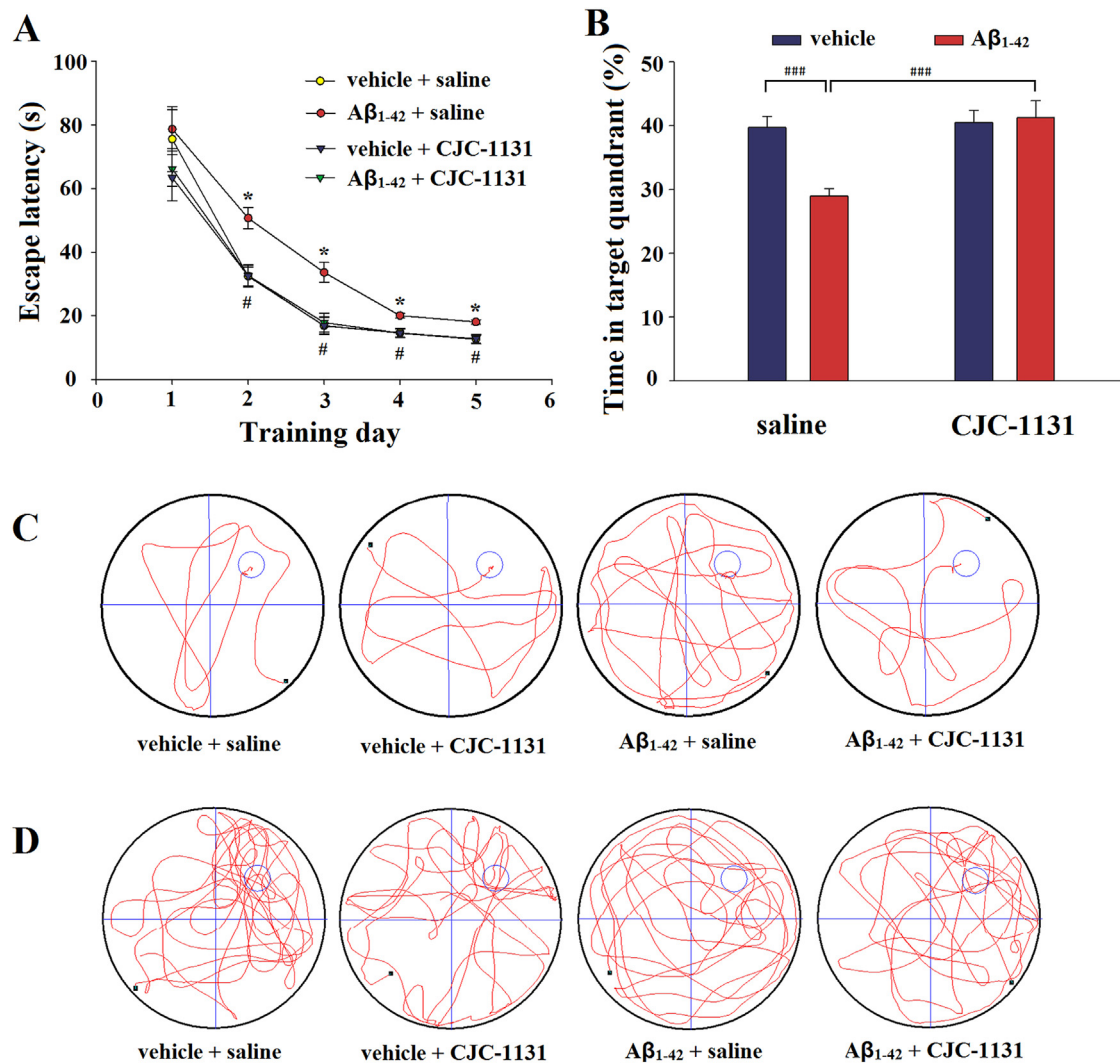


Fig. 1. The effects of Aβ₁₋₄₂ and CJC-1131 on the spatial learning and memory of rats in Morris water maze tests.

(A) Plots showing the latencies of rats to find the hidden platform over five consecutive training days in four groups. The escape latencies had a significant increase in Aβ₁₋₄₂-injected rats from the 2nd to 5th training day ($n = 10$, $^*p < 0.05$ v.s. vehicle + saline group), which was significantly reversed by CJC-1131 treatment ($n = 10$, $^{\#}p < 0.05$ v.s. Aβ₁₋₄₂ + saline group). (B) Histogram showing CJC-1131 effectively reversed Aβ₁₋₄₂-induced decrease in the swimming time in target quadrant ($n = 10$, $###p < 0.001$). Each column and point represents the mean $\pm$ SEM. (C and D) Representative original swimming traces of rats on the 3rd training day (C) and in probe trial (D) for all groups. Small circles in 1st quadrant represent the water platform, and black dots represent the locations where the rats were placed.

3. Results

3.1. CJC-1131 effectively prevented Aβ₁₋₄₂-induced impairments in spatial learning and memory of rats

As shown in Fig. 1A, the escape latencies of rats to find the hidden platform decreased during 5 consecutive training days ($F_{(4,195)} = 143.472$; $p < 0.001$). The main effect of Aβ₁₋₄₂ injection (Aβ₁₋₄₂ vs. vehicle) was significant on the escape latency ($F_{(1,38)} = 8.889$; $p < 0.01$). The main effect of CJC-1131 treatment (CJC-1131 vs. saline) was also significant ($F_{(1,38)} = 15.313$; $p < 0.001$). The interaction between Aβ₁₋₄₂ injection and CJC-1131 treatment was significant ($F_{(1,38)} = 6.567$; $p < 0.05$). The mean escape latency of rats in Aβ₁₋₄₂ injection group was significantly longer than that in vehicle injection group ($p < 0.05$), showing that the rats with Aβ₁₋₄₂ intra-hippocampal injection learned more slowly than the rats by vehicle injection. CJC-1131 treatment had no significant effect on the escape latency of rats compared with saline treatment group ($p > 0.05$), but effectively decreased the escape latency of rats

in CJC-1131 + Aβ₁₋₄₂ group, suggesting that CJC-1131 prevented against Aβ₁₋₄₂-induced deficit in learning ability.

In probe trials shown in Fig. 1B, two-way ANOVA showed that the percentage of time in the target quadrant (platform previously located) had significant main and interaction effects between Aβ₁₋₄₂ and CJC-1131 groups (Aβ₁₋₄₂ injection: $F_{(1,38)} = 8.575$; $p < 0.01$; CJC-1131 treatment: $F_{(1,38)} = 10.890$; $p < 0.01$; Aβ₁₋₄₂ by CJC-1131 interaction: $F_{(1,38)} = 6.488$; $p < 0.05$). Tukey's post hoc test showed Aβ₁₋₄₂ injection resulted in a significant decrease in the percentage of time in the target quadrant ($p < 0.001$), which was effectively reversed by CJC-1131 treatment ($p < 0.001$).

To confirm whether CJC-1131 and Aβ₁₋₄₂ affected the motor ability of rats, the average swimming speeds of rats were compared. The results showed that there was no significant difference in swimming speeds among all groups ($p > 0.05$). In addition, the results of visible platform test indicated that neither CJC-1131 nor Aβ₁₋₄₂ affected the vision of rats, since the mean escape latency in the visible platform test was nearly the same, being approximately 12.99 s in all groups.

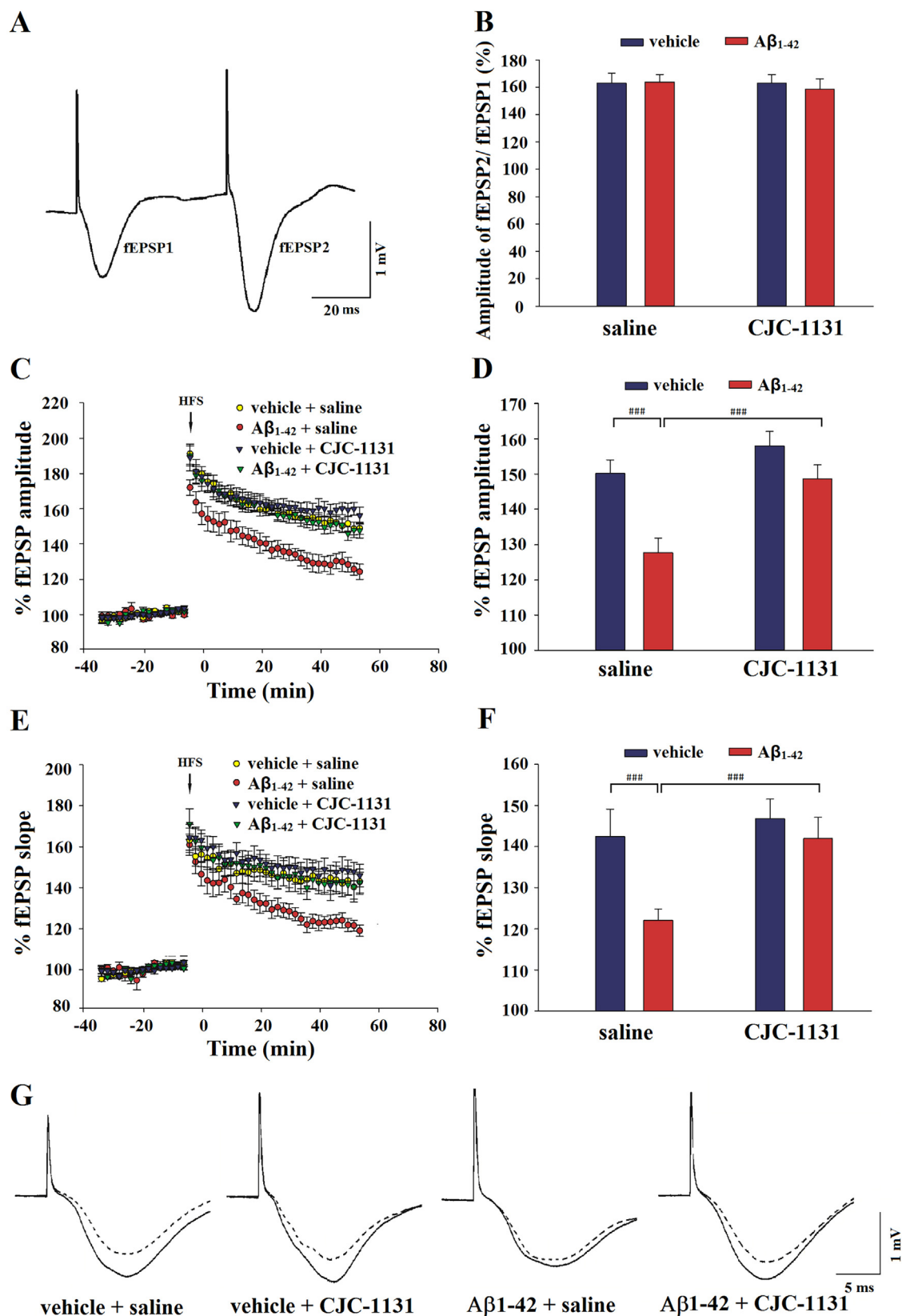


Fig. 2. The effects of A β ₁₋₄₂ and CJC-1131 on *in vivo* hippocampal LTP in the CA1 region of rats.

(A) Representative PPF trace. (B) Histograms showing neither A β ₁₋₄₂ nor CJC-1131 affected the amplitude of PPF. (C–F) Scatter plots and histograms showing CJC-1131 treatment reversed A β ₁₋₄₂-induced impairments of LTP in amplitude (C and D) and slope (E and F) ($n = 10$, $***p < 0.001$). Each column and point represents the mean $\pm$ SEM. (G) Typical fEPSP traces recorded before (dash line) and 60 min after (solid line) HFS in each group.

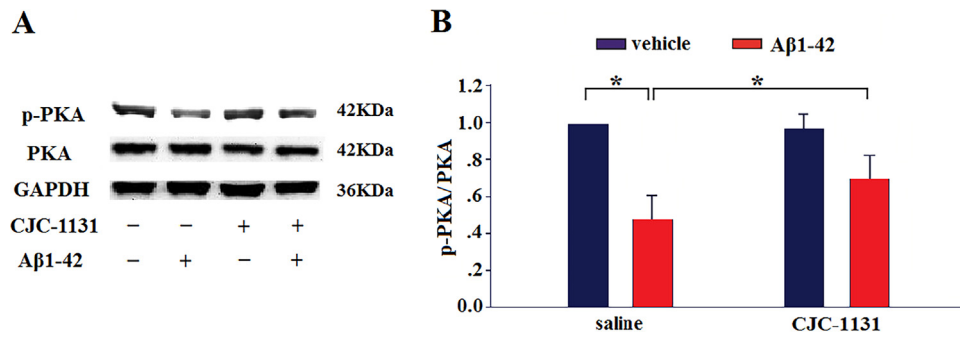


Fig. 3. CJC-1131 treatment restored the level of p-PKA in the hippocampus.

(A) The Western blot bands of p-PKA and PKA in the hippocampus, with GAPDH as the loading control. (B) Histograms showing that Aβ1-42 injection induced a significant decrease in the level of p-PKA, which was reversed by CJC-1131 treatment ($n=6$, $p<0.05$). Each column and point represents the mean $\pm$ SEM.

3.2. CJC-1131 reversed Aβ1-42-induced suppression of hippocampal LTP without affecting PPF

In view of the close relationship between spatial cognition and hippocampal synaptic plasticity, we further examined the effects of Aβ1-42 injection and CJC-1131 treatment on hippocampal synaptic plasticity. Paired-pulse facilitation (PPF) was first observed before HFS to clarify whether Aβ1-42 and CJC-1131 could affect the pre-synaptic transmitter release in the hippocampal CA1 region. Fig. 2A shows a representative PPF trace, with always higher amplitude of fEPSP2 than fEPSP1. Two-way ANOVA showed that there was no main effect on the ratio of PPF in both Aβ1-42 injection and CJC-1131 treatment (Fig. 2B), indicating that pre-synaptic neurotransmitter release was not changed by Aβ1-42 and CJC-1131. Then, we compared HFS-induced LTP by measuring the amplitude (Fig. 2C, D) and slope (Fig. 2E, F) percentages of fEPSPs. Two-way ANOVA showed that the amplitude of post-HFS had significant main and interaction effects between Aβ1-42 injection and CJC-1131 treatment groups (Aβ1-42 injection: $F(1,38)=79.876$, $p<0.001$; CJC-1131 treatment: $F(1,38)=65.105$, $p<0.001$; Aβ1-42 by CJC-1131 interaction: $F(1,38)=11.000$, $p<0.01$, Fig. 2C, D). The slope of fEPSP in all groups had similar main effects and interactions (Aβ1-42 injection: $F(1,38)=30.667$, $p<0.001$; CJC-1131 treatment: $F(1,38)=28.231$, $p<0.001$; Aβ1-42 by CJC-1131 interaction: $F(1,38)=9.809$, $p<0.01$, Fig. 2E, F). Tukey's post hoc test showed Aβ1-42 injection induced a significant decrease in the amplitude and the slope of fEPSP ($p<0.001$), while the decrease was effectively reversed by CJC-1131 treatment ($p<0.001$).

3.3. CJC-1131 restored the level of p-PKA in the hippocampus

After finishing LTP recording, we examined the level of p-PKA in the hippocampus with the same animals. Fig. 3A displayed the Western blot bands of p-PKA and PKA in the hippocampus, with GAPDH as the loading control. As shown in Fig. 3B, two-way ANOVA showed that the main effects of the p-PKA level in Aβ1-42 injection and CJC-1131 treatment were significant (Aβ1-42 injection: $F_{(1,22)}=6.499$, $p<0.05$; CJC-1131 treatment: $F_{(1,22)}=4.602$, $p<0.05$; Aβ1-42 by CJC-1131 interaction: $F_{(1,22)}=8.182$; $p<0.05$). Tukey's post hoc test showed Aβ1-42 injection induced a significant decrease in the level of p-PKA ($p<0.05$), which was effectively prevented by CJC-1131 treatment ($p<0.05$).

4. Discussion

The production and aggregation of Aβ are involved in the learning and memory deficits in AD. Not only natural Aβ such as Aβ1-40 and Aβ1-42, but also synthetic Aβ fragments like Aβ25-35 and even Aβ31-35 have strong neurotoxicity [25]. In our previous study,

Aβ31-35, similar to full length of Aβ, also induced apoptosis in cultured cortical neurons [26], formed transmembrane ion channels [27], suppressed potassium channels in neurons dissociated from the hippocampus [28,29], and impaired in vivo hippocampal LTP and spatial memory in rats [19,30].

It has been reported that Aβ desensitizes insulin receptors in cultured hippocampal neurons as well as reducing expression of the receptors [31,32]. Interestingly, these effects of Aβ can be reversed by activating GLP-1 receptor [33]. GLP-1, an insulinotropic endocrine hormone based on blood sugar concentration, can be applied to non-diabetic patients [34,35] and directly reduces endogenous Aβ levels in the mouse brain [36]. GLP-1 also promotes neurite outgrowth [37,38] and protects against oxidative injury and excitotoxic cell death [36]. In the present study, we used a new GLP-1 analogue CJC-1131, which was profoundly resistant to DPP IV degradation and has much longer half life [16]. We first confirmed in MWM test that treatment with CJC-1131 could effectively prevent against Aβ1-42-induced escape latency increase in the hidden platform test, suggesting that CJC-1131 could ameliorate Aβ1-42-induced impairment of spatial learning ability. Then, we found that CJC-1131 could effectively reversed Aβ1-42 induced decrease of swimming time in the target quadrant in probe trial, which meant that spatial memory deficit induced by Aβ1-42 could be avoided by CJC-1131. Moreover, the similar escape latency in the visible platform test and swimming speed between different groups excluded the possibility of visual or motor function deficits. We further investigated the effects of CJC-1131 on Aβ1-42-induced impairments of hippocampal synaptic plasticity. As expected, CJC-1131 prevented against Aβ1-42-induced LTP suppression, which probably explained that CJC-1131 could rescue spatial cognition impairments induced by Aβ1-42.

These results are also supported by our previous study using other GLP-1 analogs. For instance, Val(8)-GLP-1 [39], liraglutide [5] and lixisenatide [19] also reversed the suppression of LTP and the impairments of spatial learning and memory in rats. Similarly, During [38] found that the learning deficit appeared in GLP-1R-deficient mice was improved after hippocampal GLP-1R gene transfer. In addition, over expressing GLP-1R in the hippocampus also improved learning and memory of rats; application of N-AcGIP, an enzyme-resistant analogue of glucose-dependent insulinotropic polypeptide (GIP), facilitated hippocampal LTP and reversed the impairment of LTP induced by Aβ [40].

One of the possible mechanisms by which CJC-1131 prevents Aβ1-42-induced deficits in spatial cognition and synaptic plasticity could be the activation of the cAMP/PKA pathway. It is reported that Aβ can induce cytoplasmic Ca^{2+} overload [41], which results in the inhibition of AC [42], and thereby turns off a cascade reaction such as activation of cAMP and PKA. For instance, Velmurugan K found that Aβ inhibited the activity of PKA in mice cortical neurons [43];

Wang reported that A β -induced inhibition of LTP was prevented by forskolin, an activator of AC [44]; Vitolo found that A β -induced impairment of LTP was mediated through the inactivation of PKA [45]. Recently, we also found that the injection of GLP-1 agonist liraglutide could effectively increase the level of cAMP in rats brain [5]. So, we further examined the effects of GLP-1 agonist on the activity of PKA in the present study. Our results showed that A β 1-42 effectively decreased the level of p-PKA and this down-regulation could be effectively reversed by CJC-1131. It is well known that the up-regulated cAMP/PKA activity could promote gene transcription in the nucleus and finally might enhance learning and memory behavior. Therefore, we propose that the activation of cAMP/PKA pathway might be important for the neuroprotective function of GLP-1 analogs, while CJC-1131 itself could be a novel and promising candidate to protect learning and memory in neurodegenerative diseases such as AD.

Acknowledgements

This project was supported by National Science Foundation of China (31471080, 31271201), Graduate Students Outstanding Innovation Project Foundation of Shanxi Province (20143061) and National Undergraduate Innovation Program of China (201310114007, 201410114007).

References

- [1] C. Zhang, Natural compounds that modulate BACE1-processing of amyloid-beta precursor protein in Alzheimer's disease, *Discov. Med.* 14 (76) (2012) 189–197.
- [2] L. Frings, T.S. Spehl, W.A. Weber, M. Hull, P.T. Meyer, Amyloid-beta load predicts medial temporal lobe dysfunction in Alzheimer dementia, *J. Nucl. Med.* 54 (November (11)) (2013) 1909–1914.
- [3] S.S. Tang, H. Hong, L. Chen, Z.L. Mei, M.J. Ji, G.Q. Xiang, N. Li, H. Ji, Involvement of cysteinyl leukotriene receptor 1 in Abeta1-42-induced neurotoxicity in vitro and in vivo, *Neurobiol. Aging* 35 (3) (2014) 590–599.
- [4] X.H. Wang, L. Li, C. Holscher, Y.F. Pan, X.R. Chen, J.S. Qi, Val8-glucagon-like peptide-1 protects against Abeta1-40-induced impairment of hippocampal late-phase long-term potentiation and spatial learning in rats, *Neuroscience* 170 (4) (2010) 1239–1248.
- [5] W.-N. Han, C. Holscher, L. Yuan, W. Yang, X.-H. Wang, M.-N. Wu, J.-S. Qi, Liraglutide protects against amyloid- β protein-induced impairment of spatial learning and memory in rats, *Neurobiol. Aging* 34 (2) (2013) 576–588.
- [6] Z.J. Wang, W.N. Han, G.Z. Yang, L. Yuan, X.J. Liu, Q.S. Li, J.S. Qi, The neuroprotection of Rattin against amyloid beta peptide in spatial memory and synaptic plasticity of rats, *Hippocampus* 24 (1) (2014) 44–53.
- [7] P. Toro, P. Schonknecht, J. Schroder, Type II diabetes in mild cognitive impairment and Alzheimer's disease: results from a prospective population-based study in Germany, *J. Alzheimers Dis.* 16 (4) (2009) 687–691.
- [8] M.W. Strachan, R.M. Reynolds, R.E. Marioni, J.F. Price, Cognitive function, dementia and type 2 diabetes mellitus in the elderly, *Nat. Rev. Endocrinol.* 7 (2) (2011) 108–114.
- [9] M. Watve, A. Bodas, M. Diwekar, Altered autonomic inputs as a cause of pancreatic beta-cell amyloid, *Med. Hypotheses* 82 (1) (2014) 49–53.
- [10] J. Miklossy, H. Qing, A. Radenovic, A. Kis, B. Vleno, F. Laszlo, L. Miller, R.N. Martins, G. Waeber, V. Mooser, F. Bosman, K. Khalili, N. Darbinian, P.L. McGeer, Beta amyloid and hyperphosphorylated tau deposits in the pancreas in type 2 diabetes, *Neurobiol. Aging* 31 (9) (2010) 1503–1515.
- [11] Y. Chen, Z. Liang, J. Blanchard, C.L. Dai, S. Sun, M.H. Lee, I. Grundke-Iqbal, K. Iqbal, F. Liu, C.X. Gong, A non-transgenic mouse model (icv-STZ mouse) of Alzheimer's disease: similarities to and differences from the transgenic model (3xTg-AD mouse), *Mol. Neurobiol.* 47 (2) (2013) 711–725.
- [12] M.W. Schwartz, A. Sipols, S.E. Kahn, D.F. Lattemann, G.J. Taborsky Jr., R.N. Bergman, S.C. Woods, D. Porte Jr., Kinetics and specificity of insulin uptake from plasma into cerebrospinal fluid, *Am. J. Physiol.* 259 (3 Pt 1) (1990) E378–E383.
- [13] Z. Kroner, The relationship between Alzheimer's disease and diabetes: type 3 diabetes, *Altern. Med. Rev.* 14 (4) (2009) 373–379.
- [14] C.W. Chiam, J.M. Egan, Biology and therapeutic potential of GLP-1 in the treatment of diabetes, *Drug Discovery Today: Dis. Mech.* 2 (3) (2005) 295–301.
- [15] R. Leger, K. Thibaut, M. Robitaille, O. Quraishi, P. van Wyk, N. Bousquet-Gagnon, J. Carette, J.P. Castaigne, D.P. Bridon, Identification of CJC-1131-albumin bioconjugate as a stable and bioactive GLP-1(7–36) analog, *Bioorg. Med. Chem. Lett.* 14 (17) (2004) 4395–4398.
- [16] B. Lawrence, J. Dreyfus, S. Wen, P. Guivarc'h, D. Drucker, J. Castaigne, CJC-1131, a long acting GLP-1 derivative, exhibits an extended pharmacokinetic profile in healthy human volunteers, *Diabetes, AMER DIABETES ASSOC 1701 N BEAUREGARD ST, ALEXANDRIA, VA 22311-1717 USA*, (2003) A125.
- [17] R.G. Tiessen, J.P. Castaigne, J.F. Dreyfus, M. Nemansky, H.H. Kruijzinga, A.A. van Vliet, Pharmacokinetics and tolerability of a novel long-acting glucagon-like peptide-1 analog, CJC-1131, in healthy and diabetic subjects, *Int. J. Clin. Pharmacol. Ther.* 46 (9) (2008) 443–452.
- [18] L. Blonde, Clinical trial results of GLP-1-related agents: the early evidence, *Adv. Stud. Med.* 5 (10) (2005) S1074–S1078.
- [19] H.Y. Cai, C. Holscher, X.H. Yue, S.X. Zhang, X.H. Wang, F. Qiao, W. Yang, J.S. Qi, Lixisenatide rescues spatial memory and synaptic plasticity from amyloid beta protein-induced impairments in rats, *Neuroscience* 277 (2014) 6–13.
- [20] W.N. Han, C. Holscher, L. Yuan, W. Yang, X.H. Wang, M.N. Wu, J.S. Qi, Liraglutide protects against amyloid-beta protein-induced impairment of spatial learning and memory in rats, *Neurobiol. Aging* 34 (2) (2013) 576–588.
- [21] J.K. Ryu, J.G. McLarnon, Minocycline or iNOS inhibition block 3-nitrotyrosine increases and blood-brain barrier leakiness in amyloid beta-peptide-injected rat hippocampus, *Exp. Neurol.* 198 (2) (2006) 552–557.
- [22] R. Morris, Developments of a water-maze procedure for studying spatial learning in the rat, *J. Neurosci. Methods* 11 (1) (1984) 47–60.
- [23] A.V. Terry, Jr., Spatial Navigation (Water Maze) Tasks Methods of Behavior Analysis in Neuroscience, 2009 (Chapter 13).
- [24] T.V. Bliss, G.L. Collingridge, A synaptic model of memory: long-term potentiation in the hippocampus, *Nature* 361 (6407) (1993) 31–39.
- [25] L. Yuan, X.J. Liu, W.N. Han, Q.S. Li, Z.J. Wang, M.N. Wu, W. Yang, J.S. Qi, [Gly14]-Humanin protects against amyloid beta peptide-induced impairment of spatial learning and memory in rats, *Neurosci Bull Aug* 32 (4) (2016) 374–382.
- [26] X.Z. Yan, J.T. Qiao, Y. Dou, Z.D. Qiao, Beta-amyloid peptide fragment 31–35 induces apoptosis in cultured cortical neurons, *Neuroscience* 92 (1) (1999) 177–184.
- [27] J.S. Qi, J.T. Qiao, Amyloid beta-protein fragment 31–35 forms ion channels in membrane patches excised from rat hippocampal neurons, *Neuroscience* 105 (4) (2001) 845–852.
- [28] J.S. Qi, J.T. Qiao, Suppression of large conductance Ca²⁺-activated K⁺ channels by amyloid beta-protein fragment 31–35 in membrane patches excised from hippocampal neurons, *Sheng Li Xue Bao* 53 (3) (2001) 198–204.
- [29] J.S. Qi, L. Ye, J.T. Qiao, Amyloid beta-protein fragment 31–35 suppresses delayed rectifying potassium channels in membrane patches excised from hippocampal neurons in rats, *Synapse* 51 (3) (2004) 165–172.
- [30] S.F. Li, M.N. Wu, X.H. Wang, L. Yuan, D. Yang, J.S. Qi, Requirement of alpha7 nicotinic acetylcholine receptors for amyloid beta protein-induced depression of hippocampal long-term potentiation in CA1 region of rats in vivo, *Synapse* 65 (11) (2011) 1136–1143.
- [31] L. Xie, E. Helmerhorst, K. Taddei, B. Plewright, W. Van Bronswijk, R. Martins, Alzheimer's beta-amyloid peptides compete for insulin binding to the insulin receptor, *J. Neurosci.* 22 (10) (2002) (RC221).
- [32] W.Q. Zhao, F.G. De Felice, S. Fernandez, H. Chen, M.P. Lambert, M.J. Quon, G.A. Krafft, W.L. Klein, Amyloid beta oligomers induce impairment of neuronal insulin receptors, *FASEB J.* 2 (1) (2008) 246–260.
- [33] H. Gao, X. Wang, Z. Zhang, Y. Yang, J. Yang, X. Li, G. Ning, GLP-1 amplifies insulin signaling by up-regulation of IRbeta, IRS-1 and Glut4 in 3T3-L1 adipocytes, *Endocrine* 32 (1) (2007) 90–95.
- [34] D.J. Drucker, Enhancing incretin action for the treatment of type 2 diabetes, *Diabetes Care* 26 (10) (2003) 2929–2940.
- [35] B. Gallwitz, Therapies for the treatment of type 2 diabetes mellitus based on incretin action, *Minerva Endocrinol.* 31 (2) (2006) 133–147.
- [36] T. Perry, D.K. Lahiri, K. Sambamurti, D. Chen, M.P. Mattson, J.M. Egan, N.H. Greig, Glucagon-like peptide-1 decreases endogenous amyloid-beta peptide (A β) levels and protects hippocampal neurons from death induced by A β and iron, *J. Neurosci. Res.* 2 (5) (2003) 603–612.
- [37] T. Perry, D.K. Lahiri, D. Chen, J. Zhou, K.T. Shaw, J.M. Egan, N.H. Greig, A novel neurotrophic property of glucagon-like peptide 1: a promoter of nerve growth factor-mediated differentiation in PC12 cells, *J. Pharmacol. Exp. Ther.* 300 (3) (2002) 958–966.
- [38] M.J. During, L. Cao, D.S. Zuzga, J.S. Francis, H.L. Fitzsimons, X. Jiao, R.J. Bland, M. Klugmann, W.A. Banks, D.J. Drucker, C.N. Haile, Glucagon-like peptide-1 receptor is involved in learning and neuroprotection, *Nat. Med.* 9 (9) (2003) 1173–1179.
- [39] X.H. Wang, L. Li, C. Holscher, Y.F. Pan, X.R. Chen, J.S. Qi, Val8-glucagon-like peptide-1 protects against A β 1-40-induced impairment of hippocampal late-phase long-term potentiation and spatial learning in rats, *Neuroscience* 170 (4) (2010) 1239–1248.
- [40] V.A. Gault, C. Holscher, Protease-resistant glucose-dependent insulinotropic polypeptide agonists facilitate hippocampal LTP and reverse the impairment of LTP induced by beta-amyloid, *J. Neurophysiol.* 99 (4) (2008) 1590–1595.
- [41] X.H. Wang, W. Yang, C. Holscher, Z.J. Wang, H.Y. Cai, Q.S. Li, J.S. Qi, Val(8)-GLP-1 remodels synaptic activity and intracellular calcium homeostasis impaired by amyloid beta peptide in rats, *J. Neurosci. Res.* 91 (4) (2013) 568–577.
- [42] S.T. Wong, J. Athos, X.A. Figueroa, V.V. Pineda, M.L. Schaefer, C.C. Chavkin, L.J. Muglia, D.R. Storm, Calcium-stimulated adenylyl cyclase activity is critical for hippocampus-dependent long-term memory and late phase LTP, *Neuron* 23 (4) (1999) 787–798.
- [43] H. Du, L. Guo, X. Wu, A.A. Sosunov, G.M. McKhann, J.X. Chen, S.S. Yan, Cyclophilin D deficiency rescues A β -impaired PKA/CREB signaling and

- alleviates synaptic degeneration, *Biochim. Biophys. Acta* 1842 (12 Pt A) (2014) 2517–2527.
- [44] Q.W. Wang, M.J. Rowan, R. Anwyl, Inhibition of LTP by beta-amyloid is prevented by activation of beta2 adrenoceptors and stimulation of the cAMP/PKA signalling pathway, *Neurobiol. Aging* 30 (10) (2009) 1608–1613.
- [45] O.V. Vitolo, A. Sant'Angelo, V. Costanzo, F. Battaglia, O. Arancio, M. Shelanski, Amyloid beta –peptide inhibition of the PKA/CREB pathway and long-term potentiation: reversibility by drugs that enhance cAMP signaling, *Proc. Natl. Acad. Sci. U. S. A.* 99 (20) (2002) 13217–13221.